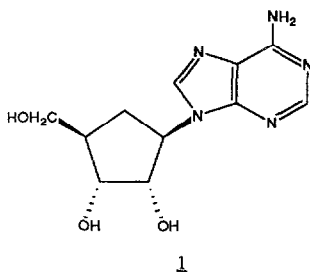


A 9-STEP ENANTIOSPECIFIC SYNTHESIS OF (-)-ARISTEROMYCIN FROM D-RIBONIC ACID γ -LACTONE

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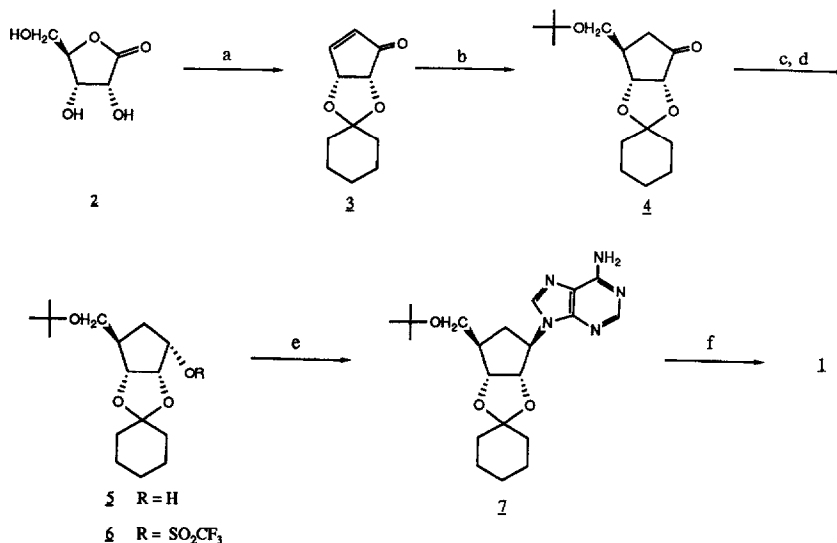
Abstract: A short, efficient, enantiospecific total synthesis of (-)-aristeromycin, which can be modified for making analogues of this carbocyclic nucleoside, is reported.

The adenosine analogue (-)-aristeromycin (**1**), isolated from *S. citricolor*,¹ is among several carbocyclic nucleoside analogues with antiviral and cytotoxic activities.² The interesting biochemical characteristics of **1**, as well as the challenge it presents to organic chemists, have prompted racemic,³ enantioselective⁴ and enantiospecific⁵ syntheses. In this communication, we report an enantiospecific total synthesis of **1**, which has the advantages, over previously reported enantiospecific syntheses, of fewer steps (9 vs. the shortest, 12) and more versatility for the synthesis of potentially bioactive analogues of **1**.



The synthesis (scheme 1) begins with D-ribonic acid γ -lactone (**2**), a useful chiral building block extolled by Bhat, et. al.⁶ Compound **2** was converted to cyclopentenone **3** in four steps as previously reported from this laboratory.⁷ This enone was treated with lithium di-(*t*-butoxymethylene)cuprate⁸ to afford the conjugate-addition product **4** in 81% purified yield.⁹ The compound was diastereomerically pure by ¹H- and ¹³C-NMR. Decoupling experiments revealed $J(\text{H}_3\text{H}_4) = 1$ Hz, corresponding to a dihedral angle of either 65° or 105° according to the Karplus equation. Molecular models illustrate that the former angle is not attainable with H₃ and H₄ in either a *cis* or *trans* relationship. The latter angle is easily attainable with these protons *trans* to

Scheme I



- a) 4 steps. See ref. 7 b) $[(\text{CH}_3)_2\text{COCH}_2]_2\text{CuLi}$, -78° to -30°C c) DIBAH, CH_2Cl_2 , 0°C , 3 h. d) $(\text{CF}_3\text{SO}_2)_2\text{O}$, pyridine, CH_2Cl_2 , 0°C , 30 min.
 e) adenine, NaH, 18-crown-6 (3 eq. each), DMF, 0°C , 18 h.¹⁰ f) $\text{CF}_3\text{COOH}/\text{H}_2\text{O}$ (2:1), 50°C , 3 h.^{13,14}

each other, but not when they are *cis*, providing evidence that the alkyl portion of the cuprate added to the least hindered face of the enone as expected.

Cyclopentanone **4** was then reduced with diisobutylaluminum hydride (DIBAH) to give alcohol **5** in 96% crude yield.¹⁰ Spectroscopic analyses indicated the compound was 92% diastereomerically pure; the two diastereomers were readily separated by silica gel chromatography (Et_2O : hexane, 1:1). Deuterium exchange of the hydroxyl proton and irradiation of $\text{H}_{5\alpha}$ and $\text{H}_{5\beta}$ revealed that $J(\text{H}_1\text{H}_2) = 5$ Hz for the major isomer and $J(\text{H}_1\text{H}_2) = 0$ Hz for the minor isomer. Again, using models and the Karplus equation, the major isomer has H_1 and H_2 in a *cis* relationship (i.e., DIBAH approached selectively from the least hindered face).

The alcohol was then converted to the triflate **6** with triflic anhydride in 95% yield.^{11,12} Displacement with sodium adenide in DMF at 0°C gave **7** in 30% yield.¹³ The chemical shift values for the purine carbons in the ^{13}C -NMR spectrum were consistent with N-9 substitution.¹⁴ No N-7 substituted isomer was detected. Mesylation¹⁵ of alcohol **5** proceeded in a quantitative crude yield, but the subsequent adenide displacement was sluggish even at 90°C . Compound **7** was deprotected by heating to 50°C in trifluoroacetic acid/water (2:1, 3h) resulting in **1** in 79% yield.^{16,17} The final compound was one diastereomer by HPLC and coeluted with an authentic sample of **1**. Spectroscopic data, optical rotation and melting point matched literature values for **1**, or, where not provided in the literature, were consistent with its structure.

Modification of the preceding sequence to provide analogues of **1** is in progress in this laboratory. Other substituents can replace the hydroxymethylene group by reacting **3** with different organocuprates. This is important in the design of more selective chemotherapeutic agents based on **1** since the hydroxymethylene group has been implicated with cytotoxic effects of carbocyclic nucleoside analogues.¹⁸

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REFERENCES AND NOTES

- 1) T. Kusaka, H. Yamamoto, M. Shibata, M. Muroi, T. Kishi and K. Mizuno. J. Antibiot., 1968, **21**, 255.
- 2) J.A. Montgomery and J.A. Secrist III. Biological Methylation and Drug Design (R.T. Borchardt, C.R. Creveling and P.M. Ueland, editors. Humana Press, Inc., Clifton, N.J., 1986), 409.
- 3) Y.F. Shealy and J.D. Clayton. J. Am. Chem. Soc., 1966, **88**, 3885; R.C. Cermak and R. Vince. Tetrahedron Lett., 1981, **22**, 2331; B.L. Kam and N.J. Oppenheimer. J. Org. Chem., 1981, **46**, 3268. G.V. Madhavan and J.C. Martin. J. Org. Chem., 1986, **51**, 1287. B.M. Trost, G.-H. Kuo and T. Benneche. J. Am. Chem. Soc., 1988, **110**, 621.
- 4) M. Arita, K. Adachi, Y. Ito, H. Sawai and M. Ohno. J. Am. Chem. Soc., 1983, **105**, 4049; Y. Arai, Y. Hayashi, M. Yamamoto, H. Takayama and T. Koizumi. Chem. Lett., 1987, **1987**, 185.
- 5) K. Tadano, M. Hoshino, S. Ogawa and T. Suami. Tetrahedron Lett., 1987, **28**, 2741. K. Tadano, M. Hoshino, S. Ogawa and T. Suami. J. Org. Chem., 1988, **53**, 1427. G.V. Madhavan and J.C. Martin. See reference 3.
- 6) K.L. Bhat, S.-Y. Chen and M.M. Joullie. Heterocycles, 1985, **23**, 691.
- 7) D.R. Borcharding, S.A. Scholtz and R.T. Borchardt. J. Org. Chem., 1987, **52**, 5457.
- 8) E.J. Corey and T.M. Eckrich. Tetrahedron Lett., 1983, **24**, 3165.
- 9) IR (KBr, cm^{-1}): 2960, 2930, 2915, 2860, 1750, 1450, 1365, 1205, 1165, 1110, 1075, 1000. $^1\text{H-NMR}$ (CDCl_3 , ppm): 1.10 (s, 9H), 1.30-1.70 (m, 10H), 2.04 (d, $J = 17$ Hz, 1H), 2.55 (dt, $J_1 = 9$ Hz, $J_2 = 2$ Hz, 1H), 2.73 (dd, $J_1 = 17$ Hz, $J_2 = 9$ Hz, 1H), 3.36 (dd, $J_1 = 9$ Hz, $J_2 = 2$ Hz, 1H), 3.55 (dd, $J_1 = 9$ Hz, $J_2 = 2$ Hz, 1H), 4.23 (d, $J = 5$ Hz, 1H), 4.62 (d, $J = 5$ Hz, 1H). $^{13}\text{C-NMR}$ (CDCl_3 , ppm): 23.6, 23.9, 25.0, 27.1, 34.0, 36.4, 37.4, 37.6, 63.1, 73.3, 78.7, 81.6, 111.5, 213.1. MS (EI, 70 eV, m/e): 282 (M^+), 226, 197, 183, 140, 98, 69, 57. M.p. = 81-83 $^\circ$ C. $[\alpha]_{\text{D}} = -154^\circ$ ($c = 0.184$, CHCl_3).
- 10) IR (neat, cm^{-1}): 3515, 2965, 2930, 2855, 1450, 1395, 1365, 1285, 1250, 1235, 1200, 1165, 1150, 1110, 1090, 1040, 1015, 955, 915, 890, 855. $^1\text{H-NMR}$ (CDCl_3 , ppm): 1.15 (s, 9H), 1.35-1.75 (m, 10H), 1.85 (m, 2H), 2.20 (m, 1H), 2.55 (d, $J = 10$ Hz, 1H, exchanged with D_2O), 3.20 (dd, $J_1 = 4$ Hz, $J_2 = 9$ Hz, 1H), 3.30 (dd, $J_1 = 4$ Hz, $J_2 = 9$ Hz, 1H), 4.20 (m, 1H), 4.45 (m, 2H). $^{13}\text{C-NMR}$ (CDCl_3 , ppm): 23.6, 24.0, 25.2, 27.4, 33.8, 36.0, 36.1, 42.2, 63.1, 72.1, 72.5, 79.2, 83.1, 111.3. MS (EI, 70 eV, m/e): 284 (M^+), 255, 241, 229, 185, 167, 113, 99, 95, 57. $[\alpha]_{\text{D}} = -13^\circ$ ($c = 0.726$, CHCl_3).
- 11) C.D. Beard, K. Baum and V. Grakauskas. J. Org. Chem., 1973, **38**, 3673.
- 12) $^1\text{H-NMR}$ (CDCl_3 , ppm): 1.10 (s, 9H), 1.25-1.75 (m, 10H), 1.95 (m, 1H), 2.25 (m, 2H), 3.20 (dd, $J_1 = 2$ Hz, $J_2 = 8$ Hz, 1H), 3.35 (dd, $J_1 = 4$ Hz, $J_2 = 8$ Hz, 1H), 4.35 (d, $J = 5$ Hz, 1H),

- 4.50 (dd, $J_1 = 5$ Hz, $J_2 = 5$ Hz, 1H), 5.25 (m, 1H). $^{13}\text{C-NMR}$ (CDCl_3 , ppm): 23.7, 23.9, 25.1, 27.2, 33.0, 34.0, 35.8, 41.4, 63.0, 73.0, 78.2, 83.2, 87.1, 112.3, 119.0 (q, CF_3).
- 13) Work up involved suction filtration, distillation of solvent, extraction between CH_2Cl_2 and water, followed by silica gel chromatography with CH_2Cl_2 : EtOH (17:3). IR (KBr, cm^{-1}): 3320, 3160, 2960, 2930, 2855, 1640, 1595, 1575, 1470, 1365, 1100, 650. $^1\text{H-NMR}$ (CDCl_3 , ppm): 1.25 (s, 9H), 1.35-1.90 (m, 10H), 2.25-2.65 (m, 3H), 3.50 (dd, $J_1 = 6$ Hz, $J_2 = 9$ Hz, 1H), 3.60 (dd, $J_1 = 4$ Hz, $J_2 = 9$ Hz, 1H), 4.65 (dd, $J_1 = 4$ Hz, $J_2 = 6$ Hz, 1H), 4.85 (m, 1H), 5.05 (dd, $J_1 = 5$ Hz, $J_2 = 6$ Hz, 1H), 6.40 (br.s., 2H), 7.95 (s, 1H), 8.35 (s, 1H). $^{13}\text{C-NMR}$ (CDCl_3 , ppm): 23.4, 23.9, 25.0, 27.4, 34.3, 34.6, 37.4, 44.0, 61.8, 62.3, 72.8, 81.2, 83.5, 113.9, 120.1, 139.4, 150.0, 152.6, 155.7. MS (EI, 70 eV, m/e): 402(M+1), 358, 344, 246, 216, 136, 57, 41. $[\alpha]_D = -32^\circ$ (c = 0.630, CHCl_3).
- 14) M.-T. Chenon, R.J. Pugmire, D.M. Grant, R.P. Panzica, and L.B. Townsend. J. Am. Chem. Soc., 1975, 97, 4627.
- 15) R.K. Crossland and K.L. Servis. J. Org. Chem., 1970, 35, 3195.
- 16) H.C. Beyerman and J.S. Bontekoe. Rec. Trav. Chim., 1962, 81, 691; S.L. Cook and J.A. Secrist III J. Am. Chem. Soc., 1979, 101, 1554.
- 17) Work up involved distillation of solvent, passage through an ion-exchange column and purification using a Hamilton PRP-1 preparative HPLC column.
- 18) M. Hasobe, J.G. McKee, D.R. Borcharding and R.T. Borchardt. Antimicrobial Agents and Chemotherapy, 1987, 31, 1849.

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